

MagicPure[®] Mouse Tissue Genomic DNA Kit

Please read the manual carefully before use.

Cat. No. EC111

Storage: The kit is at room temperature (15-25°C) for one year. Magnetic Mouse Tissue Beads at 2-8°C for one year. Avoid freezing.

Description

MagicPure[®] Mouse Tissue Genomic DNA Kit is a general-purpose kit for DNA purification of mouse tissue samples such as mouse tails and ears. Solid or liquid samples rich in impurities and inhibitors are lysed through unique lysis buffer, and DNA is specifically adsorbed with magnetic beads. The extracted DNA can be used in all kinds of molecular biology routine experiments including PCR, qPCR and so on. This kit is suitable for magnetic bar high-throughput nucleic acid extractor.

Features

- Fast extraction, high purity, high yield
- High purity, enabled by buffer solution optimized for mouse tail tissue samples and magnetic beads with high efficiency which can effectively remove inhibitive substances in downstream experiments by specific DNA adsorption.

Sample Requirements

Fresh, frozen mouse tissue. Avoid repeated freeze-thaw.

Kit Contents

Component	EC111-11 (50 rxns)
Lysis Buffer 34 (LB34)	20 ml
Proteinase K (20 mg/ml)	1 ml
Binding Buffer 34 (BB34)	10 ml
Clean Buffer 34 (CB34)	30 ml
Wash Buffer 34 (WB34)	12 ml
Elution Buffer (EB)	10 ml
Magnetic Mouse Tissue Beads	1.5 ml

Procedures

Before starting, add 100% isopropyl alcohol to BB34; add 100% ethanol to CB34 and WB 34. (refer to the table below)

Please confirm BB34 whether any crystals precipitate before use. If so, bathe at 37°C until the crystals dissolve and the solution becomes transparent.

All the magnetic separations need to be performed at room temperature. Please prepare a 70°C water bath or other heating equipment before use.

Component	EC111-11
Binding Buffer 34 (BB34)	30 ml 100% isopropyl alcohol
Clean Buffer 34 (CB34)	30 ml 100% ethanol
Wash Buffer 34 (WB34)	48 ml 100% ethanol

1. Place 10 mg of minced mouse tissue into a 1.5 ml microcentrifuge tube, and add 400 µl LB34 and 20 µl Proteinase K. Incubate at 56°C for 45 minutes, vortex 2-3 times during the incubation. (if RNA-free genomic DNA is needed, add 20 µl RNase A (Cat. No. GE01-01) to the sample).



2. Centrifuge at 12,000×g for 2 minutes, and transfer the supernatant to a new 1.5 ml microcentrifuge tube.
3. Add 500 µl BB34 and mix well. Pipette 30 µl magnetic beads into a microcentrifuge tube. Vortex for 1 minute. Incubate at room temperature for 2 minutes. Repeat 3 times, and place the microcentrifuge tube on the magnetic stand until the solution is clear.
4. Discard the supernatant carefully (avoid pipetting the beads). Add 500 µl CB34 and vortex for 2 minutes; then place the microcentrifuge tube on the magnetic stand until the solution is clear.
5. Discard the supernatant carefully (avoid pipetting the beads). Add 500 µl CB34 and vortex for 2 minutes; then place the microcentrifuge tube on the magnetic stand until the solution is clear.
6. Discard the supernatant carefully (avoid pipetting the beads). Add 500 µl WB34 and vortex for 2 minutes; then place the microcentrifuge tube on the magnetic stand until the solution is clear.
7. Discard the supernatant carefully (avoid pipetting the beads). Add 500 µl WB34 and vortex for 2 minutes; then place the microcentrifuge tube on the magnetic stand until the solution is clear.
8. Try to pipet the supernatant, air dry at room temperature for 5-10 minutes, and make ethanol fully volatilized.
9. Elution: Add 50-100 µl EB. Vortex for 30 seconds. Incubate at 65°C for 10 minutes (Mix by inversion 2-3 times during the incubation); then place the microcentrifuge tube on the magnetic stand until the solution is clear, and pipette the supernatant into a new sterile microcentrifuge tube carefully.
10. Store at -20°C.

Notes

- Avoid repeated thawing and freezing of mouse tissue to ensure quality of the extraction.
- Make sure to mix well the magnetic beads by vortexing them before use.
- Use sterile microcentrifuge tubes and pipette tips to avoid DNase contamination.
- The mouse tail tissue should be fully chopped to ensure the quality of the extraction.

FOR RESEARCH USE ONLY

