

TransTaq[®]-T DNA Polymerase

Cat. No. AP122

Concentration 5 units/μl

Storage at -20°C for two years

Description

TransTaq[®]-T DNA Polymerase is a mixture of EasyTaq[®] DNA Polymerase with a proofreading 3'-5' exonuclease. The fidelity is equal to EasyPfu DNA Polymerase. The yield is equal to that from EasyTaq[®] DNA Polymerase. It is more suitable for high fidelity TA cloning.

Highlights

- TransTaq[®]-T DNA Polymerase offers 18-fold fidelity as compared to EasyTaq[®] DNA Polymerase.
- Extension rate is about 1-2 kb/min.
- Template-independent "A" can be generated at the 3' end of the PCR product. PCR products can be directly cloned into pEASY[®]-T vectors.
- Amplification of genomic DNA fragment up to 8 kb.

Applications

- Complex templates
- TA cloning

Unit Definition

One unit of TransTaq[®]-T DNA Polymerase incorporates 10 nmol of deoxyribonucleotide into acid-precipitable material in 30 minutes at 74°C.

Quality Control

TransTaq[®]-T DNA Polymerase has passed the following quality control assays: functional absence of double- and single-strand endonuclease activity; >99% homogeneous measured by SDS-PAGE. Each batch of TransTaq[®]-T DNA Polymerase has been assayed for amplification efficiency to amplify p53 gene from 10 ng of human genomic DNA.

Storage Buffer

20 mM Tris-HCl (pH 8.0), 0.1 mM EDTA, 1 mM DTT, 100 mM KCl, 50% glycerol, stabilizers

10×TransTaq[®]-T Buffer

200 mM Tris-HCl (pH 9.0), 100 mM KCl, 100 mM (NH₄)₂SO₄, 20 mM MgSO₄, others

Kit contents

| Component | AP122-01/11 | AP122-02/12 | AP122-03/13 |
|---|---------------|---------------|---------------|
| TransTaq [®] -T DNA Polymerase | 250 U×1 | 500 U×1 | 500 U×6 |
| 10×TransTaq [®] -T Buffer | 1.2 ml ×1 | 1.2 ml ×1 | 1.2 ml ×6 |
| 2.5 mM dNTPs | - / 400 μl ×1 | - / 800 μl ×1 | - / 800 μl ×6 |
| 6×DNA Loading Buffer | 500 μl×1 | 1 ml ×1 | 1 ml ×2 |



Reaction Components

| Component | Volume | Final Concentration |
|--|---------------|---------------------|
| Template | Variable | as required |
| Forward Primer (10 μ M) | 1 μ l | 0.2 μ M |
| Reverse Primer (10 μ M) | 1 μ l | 0.2 μ M |
| 10 \times <i>TransTaq</i> [®] -T Buffer | 5 μ l | 1 \times |
| 2.5 mM dNTPs | 4 μ l | 0.2 mM |
| <i>TransTaq</i> [®] -T DNA Polymerase | 0.5-1 μ l | 2.5-5 units |
| Nuclease-free Water | Variable | - |
| Total volume | 50 μ l | - |

Thermal cycling conditions

| | | |
|---------|------------|----------------|
| 94°C | 2-5 min | } 30-35 cycles |
| 94°C | 30 sec | |
| 50-60°C | 30 sec | |
| 72°C | 1-2 kb/min | |
| 72°C | 5-10 min | |

Notes

- A final concentration of 2 mM MgSO₄ is sufficient for most targets amplification. For some targets, more Mg²⁺ may be required.
- For optimal results, we recommend to use the 100 mM MgSO₄ stock to prepare a titration from 2 mM to 4 mM (final concentration) in 0.25 mM increments.
- 0.5 μ l (2.5 units) enzyme is enough for per 50 μ l reaction. For better amplification, up to 1 μ l (5 units) enzyme can be used.

For research use only, not for clinical diagnosis.

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